

Homeopathic Medicine *Rauwolfia Serpentina* Ameliorate Blood Pressure and Oxidative Stress Parameters of Kidney by Modulating Expression of Antioxidant Enzymes in Deoxycorticosterone Acetate (DOCA)-Salt-Induced Hypertensive Rat Model

Sandeep Kumar¹, Jagneshwar Dandapat¹, Gagan Bihari Nityanand Chainy^{1*}, Akshaya Kumar Hati², Laxmikanta Nanda² and Indramani Nath³

¹Department of Biotechnology, Utkal University, Bhubaneswar, 751004, Odisha, India

²A.C.H. Medical College and Hospital, Unit III, Bhubaneswar, 751001, Odisha, India

³Department of Veterinary Surgery and Radiology, OUAT, Bhubaneswar, 751003, Odisha, India

*Corresponding author: G.B.N. Chainy, UGC Emeritus Professor, Department of Biotechnology, Utkal University, Bhubaneswar, 751004, Odisha, India, Tel/Fax: 06742567390; E-mail: chainyg@gmail.com

Received date: 14 Oct 2015; Accepted date: 11 Jan 2016; Published date: 15 Jan 2016.

Citation: Kumar S, Dandapat J, Chainy GBN, Hati AK, Nanda L, et al. (2016) Homeopathic Medicine *Rauwolfia Serpentina* Ameliorate Blood Pressure and Oxidative Stress Parameters of Kidney by Modulating Expression of Antioxidant Enzymes in Deoxycorticosterone Acetate (DOCA)-Salt-Induced Hypertensive Rat Model. J Drug Res Dev 2(1): <http://dx.doi.org/10.16966/2470-1009.111>

Copyright: © 2016 Kumar S, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

In the present study, effects of homeopathic medicine *Rauwolfia serpentina* (Q, 30c and 6c) were investigated on blood pressure, heart rate, serum biochemical parameters, oxidative stress indices and expression levels of antioxidant defence enzymes in the kidney of deoxycorticosterone acetate (DOCA)-salt-induced hypertensive uninephrectomized male rats. Administration of *R. serpentina* to hypertensive rats resulted in reduction of systolic and diastolic blood pressure. Elevated serum sodium level in hypertensive rats were reduced in response to 30c and 6c potencies of the drug. Serum potassium level was decreased in response to Q and 30c, albeit its level was increased after administration of 6c potency of the medicine. Elevated serum levels of aspartate aminotransferase (AST) and lactate dehydrogenase (LDH) of hypertensive rats were lowered in response to various potencies of the medicine. Serum glucose level was decreased and creatinine level was increased in the hypertensive rats in response to 6c potency of the medicine. Increased levels of lipid peroxidation in the kidney of hypertensive rats were restored to control value in response to all potencies of the medicine. On the other hand, elevated protein carbonyl content in the post mitochondrial fraction (PMF) of the kidney in hypertensive rats was decreased in response to 30c. However, in mitochondrial fraction (MF) it was decreased in response to the potencies, Q and 30c. Transcript level of Superoxide dismutase1 (SOD1) in the study of hypertensive rats was elevated in response to the medicine without any alteration in the level of translated products, except in case of 6c potency, where it was reduced. The transcript level of SOD2 in the kidney of hypertensive rats was decreased in response to Q potency of the medicine but its translated products were decreased in response to 30c and 6c potencies. Though SOD activity was elevated in PMF of kidney in 6c treated group, its activity was decreased in MF in response to the medicine. Although catalase (CAT) mRNA level did not change in response to the medicine, its translated products were decreased in response to 30c and 6c potencies. Kidney CAT activity of the hypertensive rats was decreased in PMF in response to all potencies of the medicine but was increased in MF in response to Q potency only. Renal glutathione peroxidase (GPx) activity of the hypertensive rats was elevated in the MF after treatment with 30c and 6c potencies of the medicine without any change in its transcript level. Glutathione S-Transferase (GST) activity of the kidney was increased in PMF of 30c treatment group. Overall findings indicate that different potencies of homeopathy medicine *R. serpentina* have the efficacy to reduce systolic blood pressure in DOCA-salt-induced hypertensive rat and also modulates serum clinical parameters and renal antioxidant defenses.

Keywords: *Rauwolfia serpentina*; DOCA; Hypertension; Blood pressure; Oxidative stress; Antioxidant

Introduction

Hypertension, the most common cardiovascular disorder, is a major risk factor for the development of stroke, coronary heart disease and renal failure [1,2]. It affects approximately 26% adult population worldwide and its prevalence is predicted to increase by 60% in 2025 [3]. Hypertension is defined as a systolic blood pressure of 140 mm Hg or higher, and a diastolic blood pressure of 90 mm Hg or higher [4]. Deoxycorticosterone acetate (DOCA)-induced hypertension is salt dependent and acts by increasing blood volume and blood pressure [5]. Prolonged administration of the DOCA with salt promotes increased concentration of aldosterone, which results in increased re-absorption of sodium ions and water from epithelial cells in the collecting ducts, causing salt and fluid retention, and it creates a volume-dependent hypertension in uninephrectomized (UNX)

rats [6,7]. Renal vascular resistance has been reported to be elevated concomitant with decrease in renal blood flow in DOCA-salt-induced hypertension, and these physiological dysfunctions are involved in the development and maintenance of hypertension [8]. Considerable evidence suggests that high blood pressure is accompanied by oxidative stress and impaired renal function in salt-sensitive hypertension [9]. DOCA-salt-induced hypertension is associated with increased reactive oxygen species (ROS) production, through NADPH oxidase, which possibly contributes to the increased blood pressure and elevated oxidative stress [10,11]. Excessive oxidative stress associated with hypertension has been documented in several hypertensive rat models, like Dahl salt sensitive [12], spontaneously hypertensive rats (SHR) [13], angiotensin II (ANG II) infused rats [14], the two-kidney one-clip Goldblatt hypertensive rats and DOCA-salt hypertension [15,10]. Furthermore, it has been observed

that these experimental models are also associated with mitochondrial alterations, which may be linked to the pathophysiology of the disease and renal damage [16].

Reserpine, the principal alkaloid of *Rauwolfia serpentina* is traditionally used for the treatment of hypertension and the first recorded reference to the application of *Rauwolfia* in case of human hypertension was in 1940 [17]. It has been observed that the roots of *R. serpentina* have antihypertensive, sedative and hypnotic properties in experimental animals [18]. It was found that prolonged use of the crude resin isolated from *R. serpentina* root produce some side effects such as sedation, depression, parkinsonism, stuffiness of the nose, suicidal tendency, and hence are not used frequently by physicians [19]. However, no such side effects have yet been observed or reported by homeopaths while using homeopathy medicine prepared from *R. serpentina* [20].

Hypertension is associated with elevated oxidative stress [21] and it is further reported that DOCA-salt-induced hypertension enhances oxidative stress through increased ROS generation [10]. Although homeopathic preparation of *R. serpentina* is traditionally used to treat hypertension in human beings by homeopathic practitioners, the present treatment regime is usually restricted to the use of mother tincture and no information is available on the use of its other potencies and their efficacy in modulating oxidative stress and antioxidant protection associated with hypertension. The present study is therefore designed to study the effect of different potencies of homeopathic medicine, *R. serpentina* (Q, 30c and 6c) on DOCA-salt-induced hypertension, serum clinical parameters, oxidative damaged products and antioxidative protection in the kidney of DOCA-salt-induced hypertensive rats.

Materials and Methods

Animals

Male Wistar rats (*Rattus norvegicus*), 120 days old and weighing 250-350 g, obtained from the National Institute of Nutrition, Hyderabad, India, and were used in the present experiment. Rats were housed in the animal room of the Department in polypropylene cages (three rats per cage) with stainless steel hopper under controlled conditions at $25 \pm 2^\circ\text{C}$ for 12:12 h light/dark cycle. The control and experimental animals were provided with food and water *ad libitum*. Entire experiment was carried out according to the guidelines of the Committee for the Purpose of Control and Supervision of Experiments on Animals, New Delhi, Gov. of India, and after the approval of experimental protocol by the Institutional Animal Ethics Committee.

Chemicals

Deoxycorticosterone acetate (DOCA) was purchased from Sigma-Aldrich Chemical Company, St. Louis, Missouri, USA. Homeopathic medicine of *R. serpentina* of different potencies (Q, 30c and 6c) prepared by Dr. Reckeweg & Co, Germany were procured from local supplier. All other chemicals used in the study were of analytical grade obtained from Sisco Research Laboratories, Mumbai, India. Serum kits were purchased from Coral clinical system, Goa, India.

Method of unilateral nephrectomy for the induction of hypertension

Rats were anesthetized with ketamine (50 mg/kg body weight) and xylazine (10 mg/kg body weight) given intramuscularly in a single dose as reported earlier [22]. The skin above the right side kidney was shaved, cleaned and applied with antiseptic. The rats were unilaterally nephrectomized via a flank incision with care being taken to preserve the adrenal gland. The kidney was freed from the surrounding tissues and gently pulled out. The renal artery and ureter were tied with silk thread and kidney was removed. The muscle and skin were sieved separately using a

sterile absorbable suture. Then rats were given antibiotic (Ampicillin 125 mg/kg body weight, intramuscular), and neomycin was sprayed on the incision area to prevent infection. After recovery the animals were used for the induction of hypertension and further experiments. Out of 30 rats, 5 rats were sham operated, and the rest were unilaterally (right kidney) nephrectomized (UNX).

Induction of hypertension and experimental design

Hypertension was induced in unilaterally nephrectomized (UNX) rats by administration of DOCA (25 mg/kg body weight given subcutaneously in 0.1 ml sesame oil/100 g body weight) twice in a week and 1% NaCl daily in drinking water *ad libitum* throughout the experimental period [22]. DOCA-salt has been reported to increase the concentration of aldosterone, which increases the reabsorption of sodium ions and water in the collecting ducts of renal tubules and thus increases the blood volume dependent hypertension [5-7]. The changes in blood pressure were monitored twice in a week by tail-cuff method (Noninvasive blood pressure NIBP). The induction of hypertension was confirmed by increased blood pressure parameters.

Experimental design for the evaluation of antihypertensive effect of *R. serpentina*

The rats were randomly divided into six groups each consisting of five animals: Sesame oil (0.1 ml/100 g body weight) was given to sham operated and unilaterally nephrectomized rats subcutaneously twice in a week.

Sham operated (Group-I) and unilaterally nephrectomized (Group-II) rats were served as control, which were provided with normal drinking water and 1% NaCl in drinking water respectively, for 65 days. UNX rats of group III-VI were given Deoxycorticosterone-acetate (DOCA) and 1% NaCl in drinking water for 35 days to induce hypertension. After 35 days of DOCA-salt treatment, hypertensive rats (Group III) were treated with the vehicle of medicine along with DOCA-salt (Group-III) and 1% NaCl, and Group IV-VI were treated with *R. serpentina* Q, 30c and 6c, along with DOCA-salt for 30 days, respectively (Scheme 1).

Blood pressure measurement

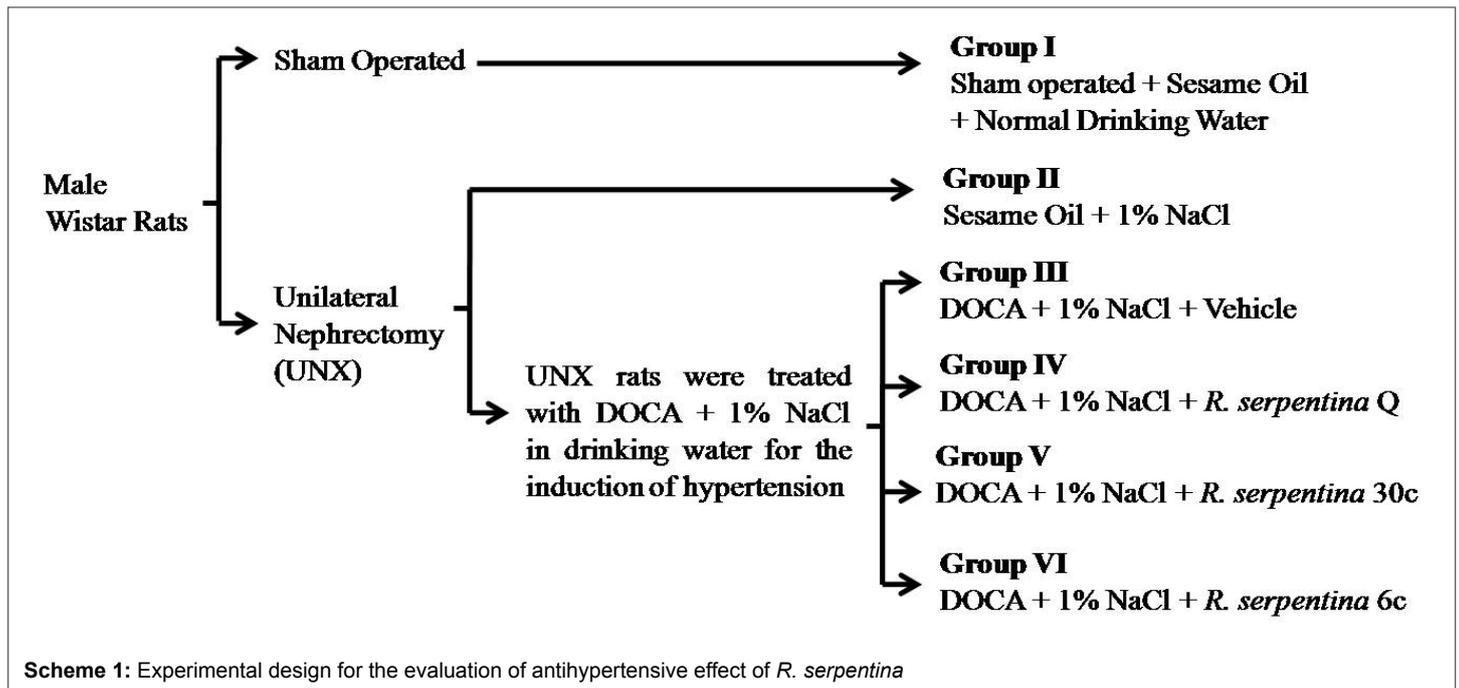
Blood pressure was measured in rats twice in a week during the study, using tail-cuff sphygmomanometer (NIBP, IITC, Life Science, USA) before starting the experiment. During blood pressure measurement, animals were kept in a heated chamber in an ambient temperature of 30-34°C for a period of 15 minutes for stabilization. From each animal 1-5 blood pressure readings were recorded and lowest three measurements were averaged. All measurements and analysis were done using computerized data acquisition system and software supplied with the tail-cuff sphygmomanometer.

Sample preparation for estimation of biochemical parameters in serum

After completion of the experiment, all animals were sacrificed under anesthesia (ketamine, 50 mg/kg body weight and xylazine, 10 mg/kg body weight) by decapitation. Immediately, trunk blood was collected from the site of decapitation in blood collection tubes with care. Then the tubes were allowed to coagulate at normal room temperature. After coagulation, tubes were centrifuged to obtain serum. Serum was stored in a deep freezer at -80°C for further biochemical analyses.

Measurement of biochemical parameters in serum

Serum electrolytes such as sodium (Na^+), potassium (K^+), chloride (Cl^-), and calcium (Ca^{2+}), serum hepatic marker enzymes like aspartate transaminase (AST), lactate dehydrogenase (LDH), alkaline phosphatase (ALP), and alanine transaminase (ALT), and serum biochemical



constituents such as, urea, urea nitrogen, uric acid, creatinine, albumin, globulin, bilirubin, total protein and glucose were measured according to the instructions of the respective kits supplied by the manufacturers.

Sample preparation for the estimation of biochemical parameters in kidney

After completion of experiment, animals were sacrificed by decapitation under anesthesia. Immediately after sacrifice, the kidneys were removed, cleaned properly and were stored in -80°C . A 10% (w/v) tissue homogenate was prepared in ice-cold homogenizing buffer (50 mM potassium phosphate buffer, pH 7.4 containing 250 mM sucrose) with a Potter-Elvehjem type motor-driven homogenizer at 4°C and subcellular fraction was carried out as described previously [23]. The crude homogenate was centrifuged at $600 \times g$ for 10 min at 4°C to pellet down the nuclei and other cell debris. The supernatant was centrifuged at $10,000 \times g$ for 20 min at 4°C to separate the mitochondria (MF) and supernatant. The resulting supernatant was called post-mitochondrial fraction (PMF). The MF was washed three times in ice cold buffer to remove cytosolic contaminants and finally dissolved in 50 mM phosphate buffer (pH 7.4). For the activity assays of antioxidant enzyme freeze-thawed MF and PMF samples were passed through Sephadex G-25. Both MF and PMF samples were used to assay SOD, GPx, GR and GST activities.

Estimation of oxidative damaged products (lipid peroxidation and protein carbonylation)

Levels of lipid peroxidation and protein carbonyl contents were measured in the PMF and MF samples and are expressed as nmoles TBARS formed/mg protein and nmoles carbonyls/mg protein, respectively [24].

Measurement of antioxidant enzymes activities

Superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx) and glutathione reductase (GR) activities were estimated as described earlier [23,24]. SOD activity was expressed as units/mg protein. CAT activity was expressed as $\mu\text{katal/mg protein}$. GPx and GR activities were expressed as nmoles of NADPH oxidized/min/mg protein. GST activity was determined following the method of Habig et al. [25] and the activity was expressed as nmoles CDNB conjugate/min/mg protein.

RNA isolation and semi-quantitative RT-PCR

Total RNA was isolated from the kidney tissue using Trizol reagent (Invitrogen Bioservices India Pvt. Ltd.). Isolated RNA samples were subjected to DNase I treatment in the presence of RNase inhibitor. Primer sequences (Table 1) of antioxidant genes were custom made by the Integrated DNA Technology, USA. G3PDH was taken as internal standard. ss-cDNA was prepared by reverse transcription of RNA (5 μg) by using 200 ng random hexamer, 20U RNase inhibitor, 1.0 mM dNTPs and 40 U of M-MuLV reverse transcriptase (Fermentas) at 37°C for 1 hour. The cDNA was then subjected to PCR at 94°C for 3 min, three steps cycle, and each cycle consisting of denaturation at 94°C for 30 s, annealing for 30 s, extension at 72°C for 1 min followed by final extension at 72°C for 5 min. The PCR mixture of 25 μl consist 1 μl RT products as template, 2.5 μl 10x buffer, 0.2 mM dNTPs, 1.5 mM MgCl_2 , 1 U *Taq* DNA polymerase (Fermentas) and 25 pmol of each primer. The PCR products were electrophoresed on 1.2% agarose gels stained with ethidium bromide. The expression levels were measured by densitometric analysis.

Preparation of tissue homogenate and western blotting

A 10% tissue homogenate was prepared in 50 mM Tris-HCl buffer, pH 7.4, containing 150 mM NaCl, 1% Triton-X-100 and 0.1% SDS and protease inhibitor to avoid protein degradation. Homogenates were centrifuged at $1,000 \times g$ for 20 min at 4°C . Thirty μg of cellular protein was resolved in 12% SDS-PAGE and transferred to PVDF membrane (0.4 μm , PALL Life Sciences) at 23 mA current for 60 min. The membrane was blocked in 5% blocking solution for 1 h at room temperature and then it was incubated with rabbit polyclonal anti-G3PDH (1:1000), anti-CAT (1:5,000), anti-SOD1 (1:5,000) and anti-SOD2 (1:2500) for 1 h at room temperature. The membrane was washed three times of 5 min each with washing solution and then incubated with HRP-conjugated anti-rabbit goat IgG (1:7500 Santa Cruz Biotechnology, Inc., USA) for 1 h at room temperature. Blot was washed with washing solution and specific immunoreactive proteins were detected with ECL kit (Santa Cruz Biotechnology, Inc., USA) in X-ray film and expression was measured with densitometry.

Gene	Accession no.	Primer Sequence (5'-3')	Annealing Temp.	Binding Position	Number of Cycles	Product size (bp)
G3PDH	DQ 403053	S-TCCCTCAAGATTGTCAGCAA AS-AGATCCACAACGGATACATT	48°C	352-371 659-640	30x	308
SOD1	NM_017150	S-GCAGAAGGCAAGCGGTGAAC AS-TAGCAGGACAGCAGATGAGT	52°C	159-178 605-586	30x	447
SOD2	BC070913	S-CTGAGGAGAGCAGCGGTCTGT AS-CTTGCCAGCGCCTCGTGGT	58°C	51-70 308-289	30x	258
CAT	BC081853	S-GCGAATGGAGAGGCAGTGTAC AS-GAGTGAGTTGTCTTCATTAGCACTG	52°C	702-722 1353-1328	30x	652
GPx1	S41066	S-CTCTCCGCGGTGGCACAGT AS-CCACCACCGGGTCGGACATAC	56.8°C	367-385 656-636	30x	290
GR	NM_053906	S-TCACTGCTCCGCACATCC AS-CTCAACACCGCCAGCGTTCTCC	54.1°C	419-436 714-693	33x	296

Table 1: Accession number, primer sequence, annealing temperature, binding position, number of PCR cycle and product sizes for amplification of antioxidant genes G3PDH, SOD1, SOD2, CAT, GPx1 and GR

Estimation of protein in tissue samples

Protein estimation in the samples was done according to the method of Bradford [26] taking BSA as standard.

Statistical analysis

Quantitation of western blot bands and PCR was done using computer assisted densitometry Image-Quant TL, Image Analysis Software v2003. Relative densities were expressed as the ratio of AOE's band/G3PDH band. Data were expressed as the mean \pm SD. The results were statistically analyzed by one-way analysis of variance (ANOVA) followed by Fisher LSD test to find out the level of significance among the mean values. The results were considered statistically significance at a level of $p \leq 0.05$.

Results

Effect of *R. serpentina* Q, 30c and 6c on body weight and different organ weight of rats (Table 2)

Body weights of rats of group I, II, III, IV, V and VI were not significantly different from each other at the end of experiment. However, relative organs weights (kidney and liver) of group II, III, IV, V and VI were higher in comparison to group I (sham-operated control). Relative weight of kidney was significantly higher in group III (1.65 fold) in comparison to group II rats. Administration of homeopathic medicine *R. serpentina* to rats of group IV, V and VI did not bring any significant change in body and organ weights in comparison to group III except for group VI, where relative kidney weight was significantly higher (1.14 fold).

Effect of *R. serpentina* Q, 30c and 6c on systolic, diastolic, mean blood pressure and heart rate of rats (Table 3)

A significant increase in systolic, diastolic, mean blood pressure and heart rate were observed in group III rats as compared to group I and group II rats. Administration of *R. serpentina* (Q, 30c and 6c) resulted in a significant decrease in the systolic blood pressure (1.15, 1.17, and 1.11 folds, respectively).

Effect of *R. serpentina* Q, 30c and 6c on serum electrolytes in rats (Table 4)

The levels of serum sodium (Na^+) increased significantly in Group III rats in comparison to group I and group II rats (1.8 and 1.2 folds, respectively). Although administration of homeopathic medicine *R. serpentina* to rats resulted in a significant decrease of serum sodium

level in group V (1.65 fold) and group VI (1.14 fold) rats, and its level remain unchanged in group IV rats as compared to group III rats. Serum potassium (K^+) level did not altered in Group III rats in comparison to group I and group II rats. A decrease in K^+ level was recorded in group IV (1.35 fold) and group V (1.35 fold) rats but its level was higher in group VI (1.53 fold) as compared to group III rats. Serum calcium level was decreased in group III rats in comparison to group I and group II rats and the decrease was 1.16 and 1.2 folds, respectively. Administration of homeopathic medicine *R. serpentina* resulted in a significant increase in serum calcium (Ca^{2+}) level in group IV (1.21 fold) and group V (1.19 fold) rats in comparison to group III rats.

Effect of *R. serpentina* Q, 30c and 6c on serum enzyme parameters (Table 5)

Serum level of AST was increased significantly in Group III rats as compared to group I and II rats (1.44 and 1.39 folds, respectively). The same trend was observed in group III rats for LDH (1.86 and 1.95 folds increased, respectively) and ALP activities (1.69 and 1.41 folds increased, respectively) as compared to group I and II rats. AST activity was decreased in response to medicine *R. serpentina* 30c (1.41 fold), however, LDH and ALP were reduced in response to all the groups receiving the medicine *R. serpentina* (Q, 30c and 6c) in comparison to DOCA-salt-induced hypertensive rats of group III. For LDH, the decrease was 1.45, 1.68 and 1.51 folds and for ALP it was 1.12, 1.32, 1.37 folds, respectively. A significant reduction in serum ALT level was noticed in group V rats (1.59 fold) in comparison to group III DOCA-salt hypertensive rats.

Effect of *R. serpentina* Q, 30c and 6c on serum biochemical parameters (Table 6)

Serum biochemical profile was not changed significantly in group III rats in comparison to group I and group II animals. Similarly not much variation was noticed in various biochemical parameters of serum of group IV, group V and group VI in comparison to group III rats. However, uric acid was decreased after supplementation of *R. serpentina* (Q, 30c, 6c) and the decrease was 1.22, 1.25 and 1.46 folds, respectively as compared to group III rats. Creatinine was decreased in group IV (2.12 fold), and glucose was decreased in group VI rats (1.63 fold) as compared to group III. Bilirubin level was decreased in group III rats as compared to group I and group II rats (1.33, 1.26 fold, respectively). After administration of *R. serpentina* bilirubin was increased in group IV, V and VI rats (1.55, 1.21 and 1.49 folds, respectively) as compared to group III rats.

Parameters		Groups					
		I	II	III	IV	V	VI
Body weight (g)	0 Day	285.0 ± 33.5	284.2 ± 19.9	320.8 ± 15.9	304.2 ± 38.3	305.8 ± 49.5	330.0 ± 36.4
	35 Day	313.0 ± 34.5	287.5 ± 23.0 ^a	347.5 ± 33.3	325.8 ± 25.6	336.7 ± 39.8	343.3 ± 50.4
	65 Day	346.7 ± 44.4	294.2 ± 28.7 ^a	350.0 ± 28.5 ^a	354.0 ± 28.2 ^a	351.7 ± 44.8 ^a	322.5 ± 46.1 ^a
Kidney Weight (g)		1.02 ± 0.14	1.30 ± 0.10 ^{ab}	1.69 ± 0.42 ^b	1.79 ± 0.31 ^b	1.77 ± 0.36 ^b	1.93 ± 0.23 ^b
KW/BW (mg/g)		3.04 ± 0.10	4.44 ± 0.28 ^d	5.13 ± 0.50 ^b	5.14 ± 0.64 ^{bc}	4.97 ± 0.50 ^{bcd}	6.03 ± 0.66 ^e
Liver Weight (g)		9.15 ± 2.72 ^a	8.14 ± 1.84 ^a	10.79 ± 1.75 ^a	12.06 ± 3.26 ^a	10.27 ± 1.72 ^a	9.77 ± 1.97 ^a
Liver/BW(mg/g)		26.5 ± 4.46 ^a	26.36 ± 4.90 ^a	29.94 ± 2.33 ^{ab}	29.85 ± 4.38 ^b	28.65 ± 2.29 ^{ab}	30.17 ± 2.65 ^{ab}

Table 2: Effect of *R. serpentina* Q, 30c and 6c potencies on body weight, kidney and liver weight in DOCA-salt induced hypertensive rats.

Group I-Sham-operated control+Normal drinking water; Group II-Nephrectomy+1% NaCl; Group III-DOCA+1% NaCl+Vehicle; Group IV-DOCA+1% NaCl+Q; Group V-DOCA+1% NaCl+30c; Group VI-DOCA+1% NaCl+6c. Data are expressed as mean ± S.D. of five animals. Data sharing superscripts of different letters differ significantly along a row at level $p \leq 0.05$.

Parameters		Groups					
		I	II	III	IV	V	VI
SBP (mm Hg)	0 Day	121.4 ± 4.3 ^a	122.8 ± 5.5 ^a	121 ± 4.5 ^a	119.4 ± 3.1 ^a	123 ± 2.6 ^a	118 ± 9.2 ^a
	35 Day	121 ± 5.1 ^a	128.2 ± 11.7 ^a	147.7 ± 6.8 ^b	150.6 ± 7.1 ^b	146.4 ± 8.1 ^b	148.6 ± 6.4 ^b
	65 Day	125.8 ± 6.9 ^a	137.2 ± 5.8 ^b	150.6 ± 6.8 ^d	130.6 ± 4.7 ^{ca}	127.8 ± 5.5 ^{ca}	134.8 ± 3.4 ^{bc}
DBP (mm Hg)	0 Day	90.2 ± 5.9 ^a	92.6 ± 6.1 ^a	89 ± 8.8 ^a	90.6 ± 6.9 ^a	90.8 ± 7.1 ^a	92 ± 5.7 ^a
	35 Day	97.0 ± 6.7 ^a	98.0 ± 10 ^{ac}	107.0 ± 13.3 ^{ab}	111.8 ± 4.3 ^b	110.0 ± 5.9 ^b	108.0 ± 3.0 ^{bc}
	65 Day	101.8 ± 7.5 ^a	108.8 ± 11.1 ^{ab}	113.0 ± 6.5 ^a	105.0 ± 4.4 ^{ab}	105.0 ± 6.8 ^{ab}	95.2 ± 12.4 ^b
MBP (mm Hg)	0 Day	77.8 ± 8.3 ^a	80.2 ± 5.4 ^a	75.2 ± 9.2 ^a	80.8 ± 7.2 ^a	83.0 ± 10.2 ^a	81.0 ± 6.7 ^a
	35 Day	84.6 ± 9.4 ^a	85.0 ± 9.9 ^a	90.2 ± 10.3 ^a	89.2 ± 6.8 ^a	87.0 ± 2.9 ^a	89.0 ± 4.4 ^a
	65 Day	85.5 ± 12.8 ^a	83 ± 16.5 ^{ab}	94.2 ± 13 ^a	89.8 ± 4.6 ^a	89.0 ± 4.4 ^a	71.2 ± 11.2 ^b
Heart Rate (BPM)	0 Day	318.0 ± 23.9	326.0 ± 2.8	312.0 ± 15.5	308.0 ± 25.1	312.8 ± 10.4	296.0 ± 15.8
	35 Day	318.6 ± 17.1 ^a	304.4 ± 24.6 ^a	409.2 ± 68.6 ^b	317.0 ± 24.9 ^a	446.4 ± 30.3 ^b	414.8 ± 38.3 ^b
	65 Day	322.0 ± 16.2 ^a	319.8 ± 14.3 ^a	452.0 ± 32.2 ^b	452.8 ± 35.4 ^b	465.0 ± 21.4 ^b	444.0 ± 20.6 ^b

Table 3: Effect of *R. serpentina* Q, 30c and 6c potencies on systolic blood pressure, diastolic blood pressure, mean blood pressure and heart rate in DOCA-salt induced hypertensive rats

Group I-Sham-operated control+Normal drinking water; Group II-Nephrectomy+1% NaCl; Group III-DOCA+1% NaCl+Vehicle; Group IV-DOCA+1% NaCl+Q; Group V-DOCA+1% NaCl+30c; Group VI-DOCA+1% NaCl+6c. Data are expressed as mean ± S.D. of five animals. Data sharing superscripts of different letters differ significantly along a row at level $p \leq 0.05$.

Parameters	Groups					
	I	II	III	IV	V	VI
Na ⁺ (mmol/l)	113.83 ± 10.22 ^{ab}	103.59 ± 9.82 ^a	123.30 ± 11.54 ^b	126.18 ± 12.10 ^b	74.64 ± 10.08 ^d	107.46 ± 4.46 ^a
K ⁺ (mmol/l)	4.84 ± 0.54 ^a	4.91 ± 0.73 ^a	4.63 ± 0.43 ^a	3.43 ± 0.35 ^b	3.42 ± 0.36 ^b	7.12 ± 1.14 ^c
Cl ⁻ (mmol/l)	89.42 ± 5.68 ^a	93.46 ± 4.21 ^a	80.03 ± 3.17 ^b	81.83 ± 3.92 ^b	80.81 ± 5.73 ^b	80.85 ± 8.06 ^b
Ca ²⁺ (mg/dl)	10.76 ± 0.8 ^a	11.04 ± 1.2 ^a	9.2 ± 1.4 ^b	11.14 ± 0.7 ^a	10.96 ± 0.7 ^a	9.82 ± 0.9 ^{ab}

Table 4: Effect of *R. serpentina* Q, 30c and 6c potencies on serum electrolytes in DOCA-salt induced hypertensive rats.

Group I-Sham-operated control+Normal drinking water; Group II-Nephrectomy+1% NaCl; Group III-DOCA+1% NaCl+Vehicle; Group IV-DOCA+1% NaCl+Q; Group V-DOCA+1% NaCl+30c; Group VI-DOCA+1% NaCl+6c. Data are expressed as mean ± S.D. of five animals. Data sharing superscripts of different letters differ significantly along a row at level $p \leq 0.05$.

Measurement of oxidatively damaged lipids and proteins (Figure 1)

The elevated levels of TBARS in the kidney of hypertensive rats were decreased in the experimental rats receiving *R. serpentina* (Q, 30c, 6c), and the decrease was 1.37, 1.42 and 1.64 folds in PMF and 1.17, 1.38 and 1.13 folds for MF, respectively, in comparison to group III rats. Administration of *R. serpentina* also resulted in a decrease in protein carbonyl levels in PMF of group V and VI rats (1.4 and 1.3 folds, respectively) and mitochondrial fraction (MF) of group IV and V rats (1.09 and 1.10 folds, respectively) as compared to group III rats.

Activity and expression levels of antioxidant enzymes

Expression of SOD (Figure 2): DOCA-salt-induced hypertensive rats treated with *R. serpentina* (Q, 30c, 6c) resulted in a significant increase in

SOD1 transcript level (1.83, 1.79 and 1.92 folds, respectively) as compared to group III rats. Decreased translated products of SOD1 in group III rats were further declined significantly in group VI (1.15 fold) i.e. in response to administration of 6c potency of *R. serpentina*. SOD activity in PMF was decreased significantly in group III rats as compared to group I and group II rats (1.63, 1.58 folds, respectively), which was further increased in group VI rats (1.91 fold) after treatment with *R. serpentina* 6c. The activity of SOD in MF was declined significantly in group IV, V and VI rats (1.91, 1.91 and 2.36 folds, respectively) in comparison to group III rats. Administration of *R. serpentina* Q resulted in the decrease of SOD2 transcript level in group IV rats (1.31 fold) and translated products in group V and VI rats (1.31 and 1.36 folds, respectively) as compared to group III rats.

Expression of CAT (Figure 3): The translated products of CAT were decreased in group V and VI (1.31 and 1.30 folds, respectively) as

Parameters	Groups					
	I	II	III	IV	V	VI
AST (U/L)	71.93 ± 12.5 ^a	74.5 ± 10.00 ^a	103.69 ± 12.54 ^{bc}	87.04 ± 14.88 ^{ac}	73.21 ± 8.10 ^a	96.26 ± 12.42 ^c
LDH (U/L)	2532.6 ± 573.1 ^{ac}	2418.8 ± 480.1 ^a	4725.2 ± 877.6 ^b	3252.8 ± 298.8 ^c	2803.8 ± 501.1 ^{ac}	3113.0 ± 646.8 ^{ac}
ALP (U/L)	62.13 ± 16.65 ^{ad}	74.11 ± 17.18 ^{acd}	105.22 ± 23.06 ^{bc}	93.308 ± 23.55 ^{cd}	79.46 ± 17.84 ^{abcd}	76.63 ± 15.92 ^d
ALT (U/L)	35.27 ± 5.00 ^a	31.47 ± 6.14 ^a	31.07 ± 5.01 ^a	31.77 ± 3.36 ^a	19.47 ± 3.86 ^b	33.87 ± 8.42 ^a

Table 5: Effect of *R. serpentina* Q, 30c and 6c potencies on serum enzymes in DOCA-salt induced hypertensive rats.

Group I-Sham-operated control+Normal drinking water; Group II-Nephrectomy+1% NaCl; Group III-DOCA+1% NaCl+Vehicle; Group IV-DOCA+1% NaCl+Q; Group V-DOCA+1% NaCl+30c; Group VI-DOCA+1% NaCl+6c. Data are expressed as mean ± S.D. of five animals. Data sharing superscripts of different letters differ significantly along a row at level $p \leq 0.05$.

Parameter	Groups					
	I	II	III	IV	V	VI
Urea (mg/dl)	26.33 ± 6.00 ^a	38.58 ± 5.24 ^b	30.09 ± 3.28 ^a	28.75 ± 6.10 ^a	30.19 ± 3.05 ^a	36.08 ± 3.53 ^{ab}
Urea Nitrogen (mg/dl)	14.19 ± 2.80 ^{ac}	18.01 ± 2.45 ^b	13.18 ± 1.48 ^{ac}	13.42 ± 2.85 ^a	14.10 ± 1.43 ^{ac}	15.88 ± 1.78 ^{bc}
Uric Acid (mg/dl)	3.60 ± 0.18 ^{ab}	3.394 ± 0.39 ^{ac}	3.85 ± 0.24 ^b	3.14 ± 0.40 ^c	3.07 ± 0.55 ^{cd}	2.62 ± 0.32 ^d
Creatinine (mg/dl)	0.55 ± 0.09 ^a	0.56 ± 0.17 ^a	0.85 ± 0.24 ^b	0.40 ± 0.05 ^a	0.64 ± 0.13 ^{ab}	1.43 ± 0.38 ^c
Albumin (g/dl)	2.772 ± 0.44 ^a	2.592 ± 0.08 ^a	2.838 ± 0.22 ^a	2.97 ± 0.26 ^a	2.67 ± 0.23 ^a	2.84 ± 0.17 ^a
Globulin (g/dl)	4.66 ± 0.38 ^{ab}	5.29 ± 0.68 ^a	4.29 ± 0.41 ^{bc}	4.51 ± 0.21 ^{bc}	3.87 ± 0.65 ^c	4.48 ± 0.52 ^{bc}
Bilirubin Total (mg/dl)	2.64 ± 0.87 ^{ab}	2.51 ± 0.55 ^{ab}	1.98 ± 0.29 ^a	3.07 ± 0.64 ^b	2.41 ± 0.48 ^{ab}	2.96 ± 0.37 ^b
Total Protein (g/dl)	7.57 ± 0.41 ^a	7.89 ± 0.70 ^a	7.71 ± 1.014 ^a	8.07 ± 0.84 ^a	7.21 ± 0.76 ^a	7.42 ± 0.35 ^a
Glucose (mg/dl)	107.27 ± 10.14 ^a	133.91 ± 15.47 ^b	81.15 ± 17.26 ^c	86.89 ± 18.28 ^{ac}	110.87 ± 13.82 ^a	49.74 ± 14.66 ^d

Table 6: Effect of *R. serpentina* Q, 30c and 6c potencies on serum biochemical parameters in DOCA-salt induced hypertensive rats.

Group I-Sham-operated control+Normal drinking water; Group II-Nephrectomy+1% NaCl; Group III-DOCA+1% NaCl+Vehicle; Group IV-DOCA+1% NaCl+Q; Group V-DOCA+1% NaCl+30c; Group VI-DOCA+1% NaCl+6c. Data are expressed as mean ± S.D. of five animals. Data sharing superscripts of different letters differ significantly along a row at level $p \leq 0.05$.

compared to group III rats. Treatment with *R. serpentina* (Q, 30c, 6c) resulted in the decrease of CAT activity in the PMF of group IV, V and VI rats (1.21, 1.42 and 1.40 folds, respectively) as compared to group III rats, however in MF it was increased in group IV rats (1.13 fold) as compared to group III rats.

Expression of GPx (Figure 4): The transcript levels of GPx1 gene and activity of GPx in PMF were found to be unaltered in DOCA-salt hypertensive rats (group III) and after treatment with *R. serpentina*. In MF, GPx activity was elevated after treatment with *R. serpentina* in group V and VI (1.15 and 1.26 folds, respectively) as compared to group III rats.

Expression of GR (Figure 5): The transcript levels of GR gene were increased in group III rats (1.39 fold) as compared to group I rats. However, its level was decreased in group IV, V and VI rats (1.18, 1.27 and 1.17 folds, respectively) as compared to group III rats. GR activity was increased in the PMF of group V and VI (1.21 and 1.16 folds, respectively) as compared to group III rats in response to *R. serpentina* Q and 6c. Elevated GR activity in MF of group III rats remains unaltered even after treatment with *R. serpentina*.

Activity of GST (Figure 5): Elevated GST activity in the PMF of kidney of hypertensive rats (group III) as compared to group I rats (control), was further increased in response to the treatment with *R. serpentina* in Q and 6c potencies (group V and VI) (1.15 and 1.10 folds, respectively). After treatment with *R. serpentina*, the GST activity in the MF was increased in group IV and VI rats (1.22 and 1.12 folds, respectively) as compared to group III rats.

Discussion

The DOCA-salt-induced hypertensive rat model is a non-genetic well established hypertension model for studying the effects of different antihypertensive compounds on hypertension [27]. In this study daily oral administration of homeopathic medicine *R. serpentina* of three potencies (Q, 30c and 6c) had resulted in significant reduction in systolic

and diastolic blood pressure parameters. The serum electrolytes such as sodium, potassium and calcium are known to play important role in blood pressure regulation [28]. DOCA-salt treated rats absorb sodium (Na^+) and water in the kidney, which causes increased circulating blood volume dependent hypertension [29]. Previous studies suggest that intracellular sodium overload and potassium depletion might be important in the pathophysiology of hypertension [30]. In the present experiment, a significant upward trend in the serum sodium levels was observed in the DOCA-salt treated rats in comparison to only unilateral nephrectomized-salt treated rats. This suggests the induction of hypertension. The reduction of sodium (in group V) and elevation of potassium (in group VI) by homeopathy medicine of *R. serpentina* suggest that this medicine depending upon its potency may be beneficial for the management of serum electrolytes level in DOCA-salt-induced hypertensive rats. A relationship between serum calcium level and high blood pressure was established by several authors. High consumption of dietary calcium is reported to prevent the rise of blood pressure [31]. A high calcium diet prevents intra myocardial vascular wall thickenings in DOCA-salt and 2-K, 1C hypertensive rats [32]. Our results are in good agreement with the results of Takata et al. [33], where a reduction in serum calcium level was recorded in hypertensive rats. Results of the present study indicate the beneficial role of homeopathy medicine *R. serpentina* in elevating serum calcium level in hypertensive rats.

Clinical diagnosis of diseases and damages to the structural integrity of the liver is commonly assessed by monitoring the serum AST, LDH, ALP and ALT activities. The increased levels of AST, ALP and LDH in serum suggest that hypertension induced by DOCA might have resulted in liver damage causing the leakage of above enzymes from the liver cells into the blood stream [34]. It is interesting to notice that the three potencies of *R. serpentina* (Q, 30c and 6c) were capable of protecting liver damages caused due to DOCA-induced hypertension.

It has been found in clinical and epidemiological studies that serum uric acid is related not only to risk of gout, but also to the risk of hypertension,

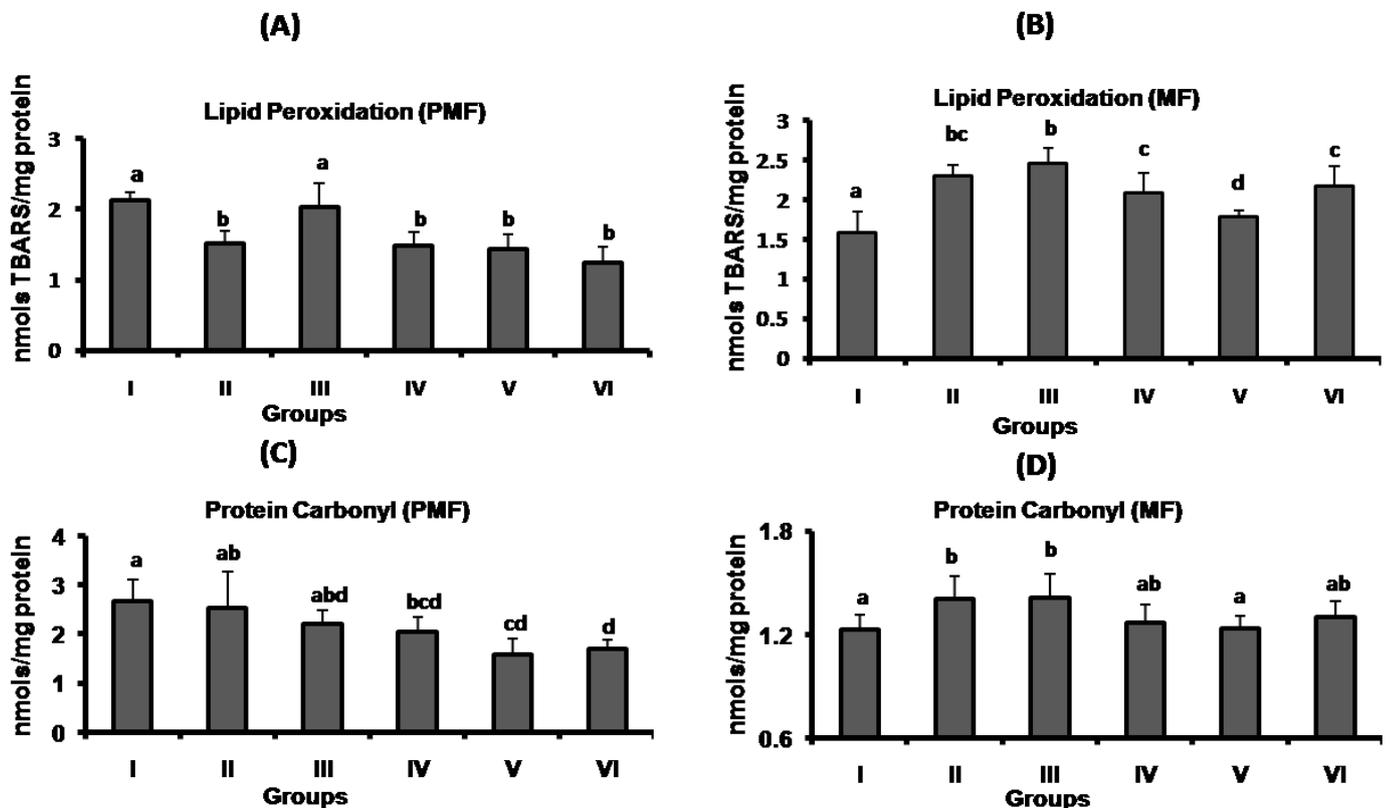


Figure 1: Effect of *R. serpentina* treatment on LPx and PC in kidneys of different groups of DOCA-salt-induced hypertensive rats. Group I (Sham operated control with normal drinking water), Group II (Nephrectomy with 1% NaCl), Group III (DOCA with 1% NaCl, Vehicle), Group IV (DOCA with 1% NaCl, *R. serpentina* Q), Group V (DOCA with 1% NaCl, *R. serpentina* 30c), Group VI (DOCA with 1% NaCl, *R. serpentina* 6c). Data are expressed as mean \pm SD of five animals. Data having different superscripts differ significantly ($p < 0.05$) from each other.

coronary heart disease, and diabetes mellitus [35]. More recently, uric acid has been proposed to have a causal role in hypertension [36]. It has been shown in many experiments that hyperuricemia causes endothelial, arteriolar and tubular interstitial damage in the kidney by increasing oxidative stress and inflammation [37,38]. Administration of medicine, *R. serpentina* considerably reduced the serum uric acid and creatinine levels in hypertensive rats. This serum uric acid may be a contributing factor to the onset of hypertension in DOCA-salt-induced hypertensive rats [39].

Bilirubin acts as a potent physiological antioxidant which can protect from coronary artery disease [40]. Serum bilirubin has been consistently shown to be inversely related with cardiovascular disease. Recent studies showed that serum bilirubin is associated with cardiovascular disease related factors such as diabetes, body mass index and metabolic syndrome [41]. Increased serum bilirubin levels in *R. serpentina* treated groups suggest that the medicine may give additional protection to heart during hypertension state.

Serum glucose level was reduced in hypertensive rats in response to 6c potency of *R. serpentina* which is similar to the earlier report of Azmi et al. [42]. The authors have reported that methanol extracts of *R. serpentina* exhibit hypoglycemic activity in alloxan-induced diabetic rat model. Such hypoglycemic effect of 6c potency of *R. serpentina* might be either due to reduced glucose absorption in intestine or due to augmented uptake of glucose by different tissues from blood.

In this study we have found that the administration of DOCA-salt

increases blood pressure parameters which are consistent with the previous studies of Kwak et al. [43] and Chen et al. [44]. ROS are generated as normal products of cellular metabolism. Free radical mediated lipid peroxidation has been implicated in the pathogenesis of many diseases such as cardiovascular diseases, cancer, neurological disorders and aging [45,46]. Increased levels of LPx and PC in the kidney of DOCA-salt treated rats suggest elevated level of oxidative stress in hypertension. It has been reported that hypertension is associated with hyperlipidemia, which promotes lipid peroxidation and enhances oxidative stress [47]. Products of lipid peroxidation such as 4-hydroxynonenal, malondialdehyde etc. binds covalently to amino acid residues of proteins and causes oxidative modification of proteins in kidney [48]. In this study the increased level of lipid peroxidation and protein carbonyl content in the tissue of DOCA-salt hypertensive rats may be due to increased oxidative stress. The reduction in the LPx and protein carbonyl content in uninephrectomized DOCA-salt treated rats after administration of *R. serpentina* indicates the antioxidative potential of the drug against the hypertension.

Over production or declined elimination of ROS in aerobic cells can lead to their accumulation causing oxidative stress. So all oxygen metabolizing cells have evolved antioxidant defence mechanisms to protect from oxidative damage by the ROS. It has been reported that vascular superoxide formation is enhanced in DOCA-salt hypertensive rats [49] by nicotinamide adenine dinucleotide phosphate (NADPH) oxidase [10]. Long-term administration of superoxide radical scavenger, tempol or NADPH-oxidase inhibitor, apocynin decreased systolic blood pressure

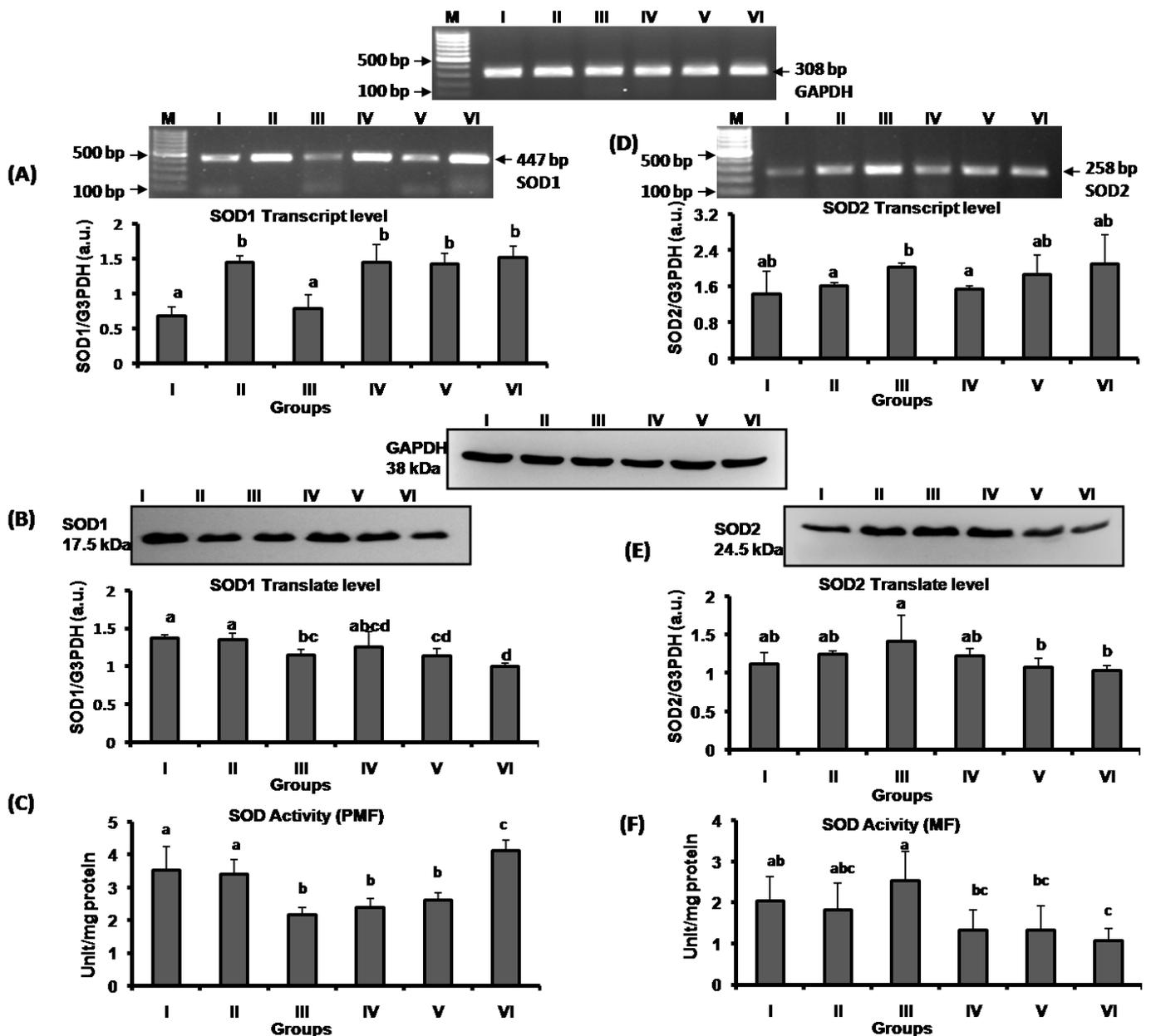


Figure 2: Effect of *R. serpentina* treatment on expression of renal antioxidant enzyme SOD1 at the level of mRNA (A), translated product (B) and enzyme activity in PMF (C) of control, sham, hypertensive and *R. serpentina* treated rats and expression of SOD2 at the mRNA level (D), translated product (E) and enzyme activity in MF (F). Group I (Sham operated control with normal drinking water), Group II (Nephrectomy with 1% NaCl), Group III (DOCA with 1% NaCl, Vehicle), Group IV (DOCA with 1% NaCl, *R. serpentina* Q), Group V (DOCA with 1% NaCl, *R. serpentina* 30c), Group VI (DOCA with 1% NaCl, *R. serpentina* 6c). Quantitative representations of SOD1/G3PDH and SOD2/G3PDH of mRNA and proteins obtained by densitometric analyses and expressed as arbitrary units (a.u.). Data are expressed as means \pm S.D. of five animals. Data having different superscripts differ significantly ($p \leq 0.05$) from each other.

and reduced superoxide radical generation in DOCA-salt hypertensive rats [10]. Role of antioxidants in scavenging ROS and protecting the cells from oxidative damage in hypertension condition is well established [50]. Enzymatic antioxidants present in the body include SOD, catalase and glutathione peroxidase, which act as first line of defence against ROS by catalyzing their conversion to less reactive substances [51]. SOD enzyme effectively dismutates highly reactive and toxic superoxide radicals into H_2O_2 [50]. Catalase catalyzes harmful hydrogen peroxide molecules and converts it into water and oxygen. On the other hand, GPx catalyzes

the reduction of hydrogen peroxide and hydroperoxide molecules to nontoxic products. Our study indicates that different potencies of the drug differentially regulate the expression of antioxidant defence enzymes in activity, transcript and translate levels. The transcript levels of SOD2 were more elevated in hypertensive groups than control, and medicine treatment reduced the mRNA level and activity in the mitochondrial fraction. The same trend was observed for the translated products of SOD2. In contrast, activities of SOD1 exhibited the reverse trend at both transcript and translate levels. These findings may be due to the fact that

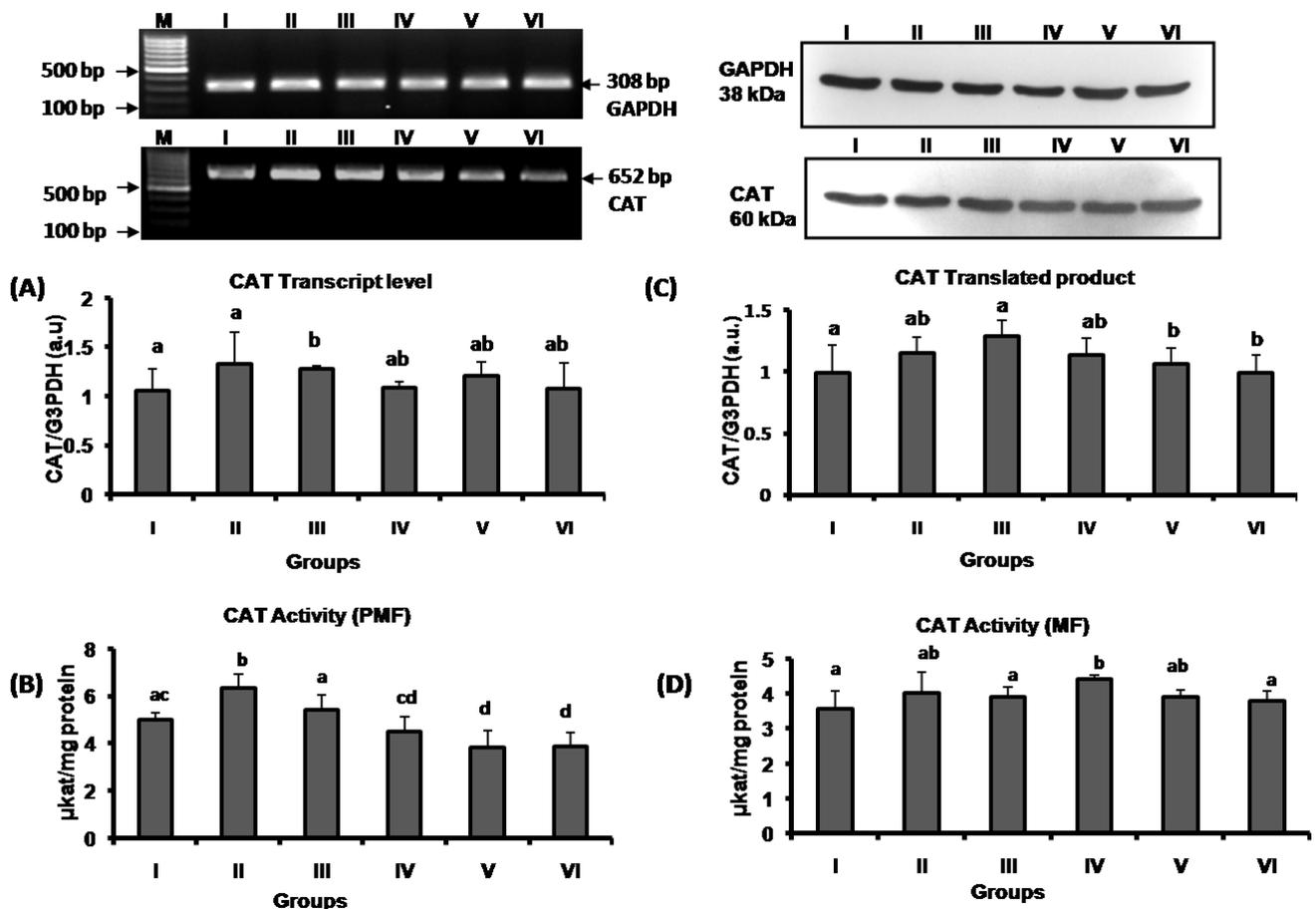


Figure 3: Effect of *R. serpentina* treatment on expression of renal antioxidant enzyme CAT at the level of mRNA (A), enzyme activity in PMF (B), translated product (C) and enzyme activity in MF (D) of control, sham, hypertensive and *R. serpentina* treated rats. Group I (Sham operated control with normal drinking water), Group II (Nephrectomy with 1% NaCl), Group III (DOCA with 1% NaCl, Vehicle), Group IV (DOCA with 1% NaCl, *R. serpentina* Q), Group V (DOCA with 1% NaCl, *R. serpentina* 30c), Group VI (DOCA with 1% NaCl, *R. serpentina* 6c). Quantitative representations of CAT/G3PDH of mRNA and proteins obtained by densitometric analyses and expressed as arbitrary units (a.u.). Data are expressed as means \pm S.D. of five animals. Data having different superscripts differ significantly ($p \leq 0.05$) from each other.

SOD1 is active in cytoplasm and SOD2 is active in mitochondria and their promoter regions differ greatly between the two enzymes. Our findings are in line with the study of Chaves et al. [52].

It is a well established fact that mitochondrial respiration is accelerated due to DOCA-salt [53]. Enhancement of cellular respiration is generally associated with increased production of superoxide radicals at the site of ubiquinone [53]. Our results suggest that the alteration of antioxidant enzymes in response to *R. serpentina* treatment to DOCA-salt treated rats is modulated through distinct regulatory pathways. *R. serpentina* protect the rat kidney against oxidative stress induced by DOCA-salt hypertension. However, different potencies of *R. serpentina* differently exhibit their effects on oxidative stress parameters and antioxidant defence system in hypertensive rats.

Overall findings of our study indicate that administration of *R. serpentina* (Q, 30c and 6c) reduces blood pressure, as evident from reduced blood pressure in hypertensive rats. We, for the first time observed that homeopathic medicine *R. serpentina* (Q, 30c and 6c) can decrease hypertension effectively and Q potency was the most effective in the regulation of blood pressure. It has not shown any drug toxic effect on the organs, which is evidenced by the serum biochemical parameters.

R. serpentina also exhibited antioxidant potential in this study as it could attenuate oxidative stress indices in the kidney of DOCA-salt-induced hypertensive rats. Thus *R. serpentina* has both antihypertensive and antioxidant properties, as evident from considerable decrease in the blood pressure, oxidative stress parameters and modulation of enzymatic antioxidants in response to drug treatment. These results provide an important pharmacological basis for the use of this homeopathically prepared medicine to prevent the hypertension and associated oxidative stress.

Acknowledgement

The work was generously funded under Extra Mural Research project of Central Council for Research in Homeopathy by Department of AYUSH, Ministry of Health and Family Welfare, Govt. of India, New Delhi (F. No. 23-94/2006-07/CCRH. Tech/EMR/1330. Dt. 10.01.2008). We are highly thankful to the Principal, Dr. A.C.H. Medical College and Hospital, Bhubaneswar, India and to the Head, Post Graduate Department of Biotechnology, Utkal University, Bhubaneswar, India to allow for conducting the study and for providing laboratory facility. Authors also acknowledge the support of DST-PUSRE grant (Gov. of India) to the Department of Biotechnology, Utkal University, Bhubaneswar.

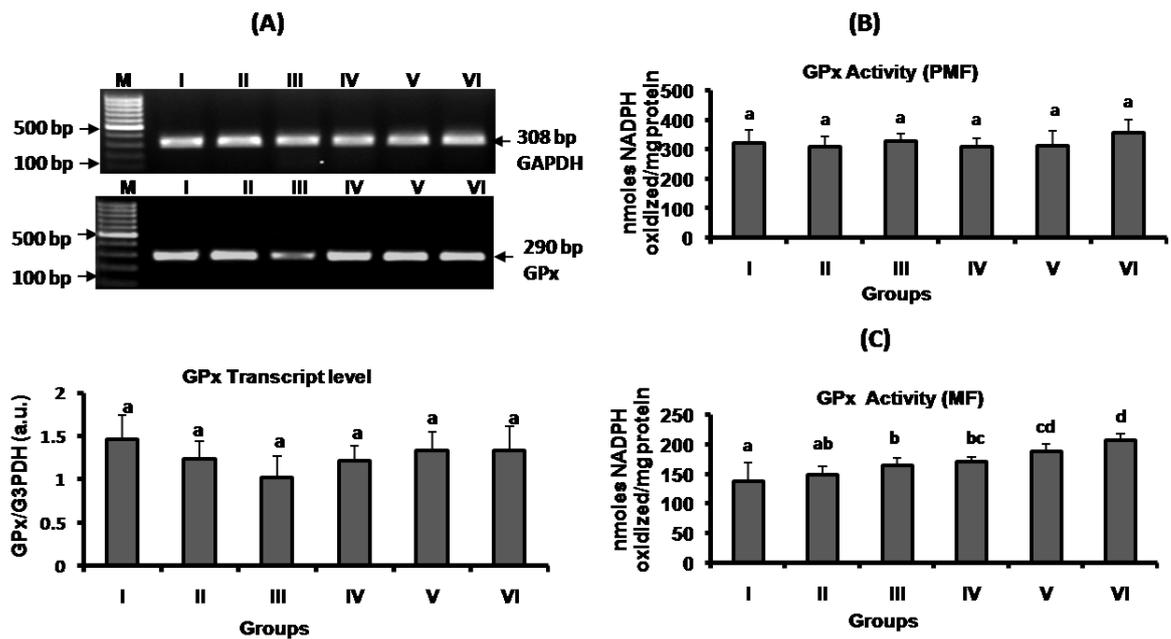


Figure 4: Effect of *R. serpentina* treatment on expression of renal antioxidant enzyme GPx at the level of mRNA (A), enzyme activity in PMF (B), and enzyme activity in MF (C) of control, sham, hypertensive and *R. serpentina* treated rats. Group I (Sham operated control with normal drinking water), Group II (Nephrectomy with 1% NaCl), Group III (DOCA with 1% NaCl, Vehicle), Group IV (DOCA with 1% NaCl, *R. serpentina* Q), Group V (DOCA with 1% NaCl, *R. serpentina* 30c), Group VI (DOCA with 1% NaCl, *R. serpentina* 6c). Quantitative representations of GPx/G3PDH of mRNA obtained by densitometric analyses and expressed as arbitrary units (a.u.). Data are expressed as means \pm S.D. of five animals. Data having different superscripts differ significantly ($p \leq 0.05$) from each other.

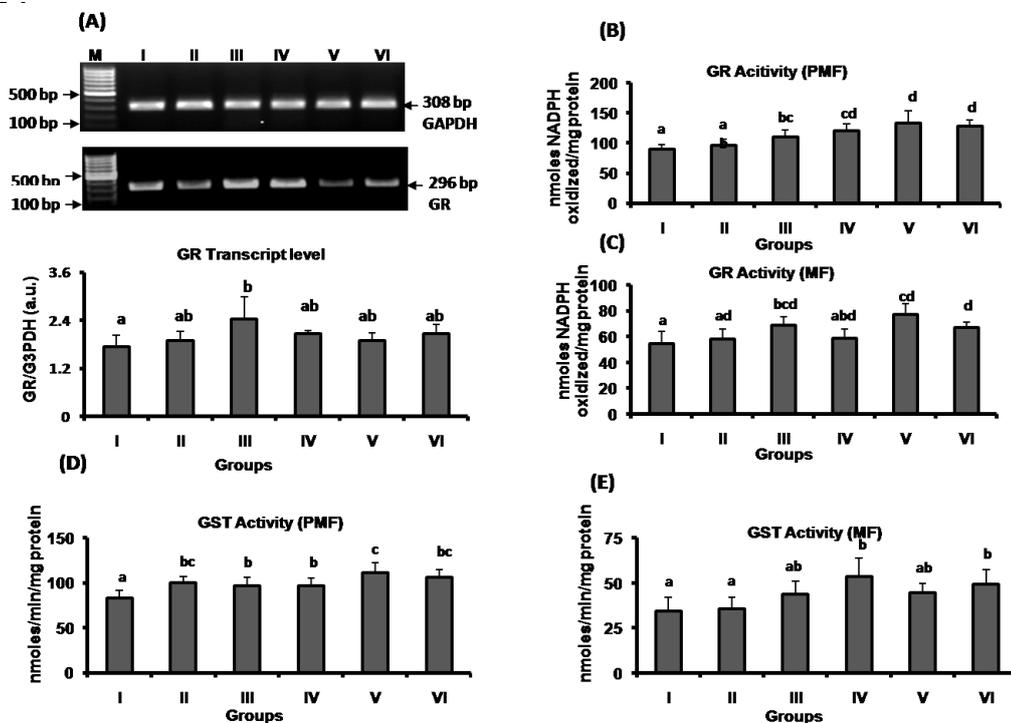


Figure 5: Effect of *R. serpentina* treatment on expression of renal antioxidant enzyme GR at the level of mRNA (A), enzyme activity in PMF (B), and enzyme activity in MF (C); GST enzyme activity in PMF (D) and MF (E) of control, sham, hypertensive and *R. serpentina* treated rats. Group I (Sham operated control with normal drinking water), Group II (Nephrectomy with 1% NaCl), Group III (DOCA with 1% NaCl, Vehicle), Group IV (DOCA with 1% NaCl, *R. serpentina* Q), Group V (DOCA with 1% NaCl, *R. serpentina* 30c), Group VI (DOCA with 1% NaCl, *R. serpentina* 6c). Quantitative representations of GR/G3PDH of mRNA obtained by densitometric analyses and expressed as arbitrary units (a.u.). Data are expressed as means \pm S.D. of five animals. Data having different superscripts differ significantly ($p \leq 0.05$) from each other.

References

1. Bittar N (1995) Maintaining long-term control of blood pressure: the role of improved compliance. *Clin Cardiol* 18: III 12-16.
2. Briones AM, Touyz RM (2010) Oxidative stress and hypertension: current concepts. *Curr Hypertens Rep* 12: 135-142.
3. Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, et al. (2005) Global burden of hypertension: analysis of worldwide data. *Lancet* 365: 217-223.
4. Peters PG, Alessio HM, Hagerman AE, Ashton T, Nagy S, et al. (2006) Short-term isometric exercise reduces systolic blood pressure in hypertensive adults: possible role of reactive oxygen species. *Int J Cardiol* 110: 199-205.
5. Dahl LK (2005) Possible role of salt intake in the development of essential hypertension. *Int J Epidemiol* 34: 967-972.
6. Ortiz PA, Garvin JL (2001) Intrarenal transport and vasoactive substances in hypertension. *Hypertension* 38: 621-624.
7. Tomaschitz A, Pilz S, Ritz E, Obermayer-Pietsch B, Pieber TR (2010) Aldosterone and arterial hypertension. *Nat Rev Endocrinol* 6: 83-93.
8. Huang M, Hester RL, Coleman TG, Smith MJ, Guyton AC (1992) Development of hypertension in animals with reduced total peripheral resistance. *Hypertension* 20: 828-833.
9. Seifi B, Kadkhodae M, Karimian SM, Zahmatkesh M, Xu J, et al. (2010) Evaluation of renal oxidative stress in the development of DOCA-salt induced hypertension and its renal damage. *Clin Exp Hypertens* 32: 90-97.
10. Beswick RA, Zhang H, Marable D, Catravas JD, Hill WD, et al. (2001) Long-term antioxidant administration attenuates mineralocorticoid hypertension and renal inflammatory response. *Hypertension* 37: 781-786.
11. Roberts CK, Barnard RJ, Sindhu RK, Jurczak M, Ehdaie A, et al. (2006) Oxidative stress and dysregulation of NAD(P)H oxidase and antioxidant enzymes in diet-induced metabolic syndrome. *Metabolism* 55: 928-934.
12. Meng S, Roberts LJ, Cason GW, Curry TS, Manning RD Jr. (2002) Superoxide dismutase and oxidative stress in Dahl salt-sensitive and resistant rats. *Am J Physiol Regul Integr Comp Physiol* 283: R732-R738.
13. Welch WJ, Wilcox CS (2001) AT1 receptor antagonist combats oxidative stress and restores nitric oxide signaling in the SHR. *Kidney Int* 59: 1257-1263.
14. Reckelhoff JF, Zhang H, Srivastava K, Roberts LJ 2nd, Morrow JD, et al. (2000) Subpressor doses of angiotensin II increase plasma F2-isoprostanes in rats. *Hypertension* 35: 476-479.
15. Xu H, Fink GD, Galligan JJ (2004) Tempol lowers blood pressure and sympathetic nerve activity but not vascular O₂ in DOCA-salt rats. *Hypertension* 43: 329-334.
16. Zuniga-Toala A, Tapia E, Zazueta C, Correa F, Lucia Z, et al. (2012) Nordihydroguaiaretic acid pretreatment prevents ischemia and reperfusion induced renal injury, oxidant stress and mitochondrial alterations. *J Med Plants Res* 6: 2938-2947.
17. Vakil RJ (1940) Hypertension. *Med Bull* 8: 495-503.
18. Chopra RN, Gupta JC, Mukherjee SN (1933) The pharmacological action of an alkaloid obtained from *Rauwolfia serpentina* Benth. A preliminary note. *Indian J Med Res* 21: 261-271.
19. Gupta JC, Kahali BS, Dutt A (1944) The hypnotic action of resin fraction isolated from root of *Rauwolfia serpentina* Benth. Obtained from Dehradun. *Indian J Med Res* 32: 183-188.
20. Rastogi DP, Baig H (1996) *Rauwolfia serpentina* (Aqua): A New Approach in the Treatment of Hypertension in Homoeopathy. *CCRH* 18: 22-24.
21. Somers MJ, Mavromatis K, Galis ZS, Harrison DG (2000) Vascular superoxide production and vasomotor function in hypertension induced by deoxycorticosterone acetate-salt. *Circulation* 101: 1722-1728.
22. Dallemagne C, Ooi SY, Bown L, Gobe G, Endre Z (2000) Renal impairment in deoxycorticosterone acetate-salt hypertensive rats. *Nephrology* 5: 277-284.
23. Das K, Samanta L, Chainy GBN (2000) A modified spectrophotometric assay of superoxide dismutase using nitrite formation of superoxide radicals. *Indian J Biochem Biophys* 37: 201-201.
24. Jena S, Chainy GBN, Dandapat J (2012) Expression of antioxidant genes in renal cortex of PTU-induced hypothyroid rats: effect of vitamin E and curcumin. *Mol Biol Rep* 39: 1193-1203.
25. Habig WH, Pabst MJ, Jakoby WB (1974) Glutathione S-transferases. The first enzymatic step in mercapturic acid formation. *J Biol Chem* 249: 7130-7139.
26. Bradford MM (1976) A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. *Anal Biochem* 72: 248-254.
27. Iyer A, Chan V, Brown L (2010) The DOCA-salt hypertensive rat as a model of cardiovascular oxidative and inflammatory stress. *Curr Cardiol Rev* 6: 291-297.
28. Karppanen H (1991) Minerals and blood pressure. *Ann Med* 23: 299-305.
29. Leiba A, Vald A, Peleg E, Shamiss A, Grossman E (2005) Does dietary recall adequately assess sodium, potassium and calcium intake in hypertensive patients? *Nutrition* 21: 462-466.
30. Krishna GG, Kapoor SC (1991) Potassium depletion exacerbates essential hypertension. *Ann Intern Med* 115: 77-83.
31. Perry PA (1995) Effect of supplemental dietary calcium on the development of DOCA-salt hypertension in weanling rats. *West J Nurs Res* 17: 63-75.
32. Liu DT, Birchall I, Hewitson T, Kincaid-Smith P, Whitworth JA (1994) Effect of dietary calcium on the development of hypertension and hypertensive vascular lesions in DOCA-salt and two-kidney, one-clip hypertensive rats. *J Hypertens* 12: 145-153.
33. Takata Y, Yamashita Y, Takishita S, Fujishima M (1990) DOCA-salt treatment decreases serum total calcium but not ionized calcium concentration in rats. *Am J Hypertens* 3: 237-239.
34. Concepcion Navarro M, Pilar Montilla M, Martin A, Jimenez J, Pilar Utrilla M (1993) Free radicals scavenger and antihepatotoxic activity of Rosmarinus. *Planta Med* 59: 312-314.
35. Li Y, Stamler J, Xiao Z, Folsom A, Tao S, et al. (1997) Serum uric acid and its correlates in Chinese adult populations, urban and rural, of Beijing. The PRC-USA Collaborative Study in Cardiovascular and Cardiopulmonary Epidemiology. *Int J Epidemiol* 26: 288-296.
36. Feig DI, Madero M, Jalal DI, Sanchez-Lozada LG, Johnson RJ (2013) Uric acid and the origins of hypertension. *J Pediatr* 162: 896-902.
37. Sanchez-Lozada LG, Soto V, Tapia E, Avila-Casado C, Sautin YY, et al. (2008) Role of oxidative stress in the renal abnormalities induced by experimental hyperuricemia. *Am J Physiol Renal Physiol* 295: F1134-F1141.
38. Kosugi T, Nakayama T, Heinig M, Zhang L, Yuzawa Y, et al. (2009) Effect of lowering uric acid on renal disease in the type 2 diabetic db/db mice. *Am J Physiol Renal Physiol* 297: F481-F488.
39. Seifi B, Kadkhodae M, Zahmatkesh M (2011) Effect of vitamin E therapy on serum uric acid in DOCA-salt treated rats. *Acta Physiol Hung* 98: 214-220.
40. Mayer M (2000) Association of serum bilirubin concentration with risk of coronary artery disease. *Clin Chem* 46: 1723-1727.

Citation: Kumar S, Dandapat J, Chainy GBN, Hati AK, Nanda L, et al. (2016) Homeopathic Medicine *Rauwolfia Serpentina* Ameliorate Blood Pressure and Oxidative Stress Parameters of Kidney by Modulating Expression of Antioxidant Enzymes in Deoxycorticosterone Acetate (DOCA)-Salt-Induced Hypertensive Rat Model. *J Drug Res Dev* 2(1): <http://dx.doi.org/10.16966/2470-1009.111>

41. Lin JP, Vitek L, Schwertner HA (2010) Serum bilirubin and genes controlling bilirubin concentrations as biomarkers for cardiovascular disease. *Clin Chem* 56: 1535-1543.
42. Azmi MB, Qureshi SA (2012) Methanolic root extract of *Rauwolfia serpentina* improves the glucose tolerance in Wister mice. *J Food Drug Anal* 20: 484-488.
43. Kwak CJ, Kubo E, Fujji K, Nishimura Y, Kobuchi S, et al. (2009) Antihypertensive effect of French maritime pine bark extract (Flavangenol): possible involvement of endothelial nitric oxide-dependent vasorelaxation. *J Hypertens* 27: 92-101.
44. Chen YY, Ji W, Du JR, Yu DK, He Y, et al. (2010) Preventive effects of low molecular mass potassium alginate extracted from brown algae on DOCA salt-induced hypertension in rats. *Biomed Pharmacother* 64: 291-295.
45. Leonarduzzi G, Gamba P, Gargiulo S, Biasi F, Poli G (2012) Inflammation-related gene expression by lipid oxidation-derived products in the progression of atherosclerosis. *Free Radic Biol Med* 52: 19-34.
46. Negre-Salvayre A, Auge N, Ayala V, Basaga H, Bosada J, et al. (2010) Pathological aspects of lipid peroxidation. *Free Radic Res* 44: 1125-1171.
47. Sutken E, Inal M, Ozdemir F (2006) Effects of vitamin E and gemfibrozil on lipid profiles, lipid peroxidation and antioxidant status in the elderly and young hyperlipidemic subjects. *Saudi Med J* 27: 453-459.
48. Zarkovic N (2003) 4-Hydroxyneonal as a bioactive marker of pathophysiological processes. *Mol Aspects Med* 24: 281-291.
49. Fridovich I (1999) Fundamental aspects of reactive oxygen species, or what's the matter with oxygen. *Ann N Y Acad Sci* 893: 13-18.
50. Mates JM (2000) Effects of antioxidant enzymes in the molecular control of reactive oxygen species toxicology. *Toxicology* 153: 83-104.
51. Veil EC, Benkirane K, Javeshghani D, Touyz RM, Schiffrin EL (2008) Xanthine oxidase and mitochondria contribute to vascular superoxide anion generation in DOCA-salt hypertensive rats. *Am J Physiol, Heart Circ Physiol* 295: H281-H288.
52. Chaves FJ, Mansego ML, Blesa S, Gonzalez-Albert V, Jimenez J, et al. (2007) Inadequate cytoplasmic antioxidant enzymes response contributes to the oxidative stress in human hypertension. *Am J Hypertens* 20: 62-69.
53. Venditti P, Pamplona R, Portero-Otin, M, De Rosa R, Di Meo S (2006) Effects of experimental and cold exposure induced hyperthyroidism on H₂O₂ production and susceptibility to oxidative stress of rat liver mitochondria. *Arch Biochem Biophys* 447: 11-22.